

## *n*-ALKANES AND $\omega$ -HYDROXYALKANOIC ACIDS FROM THE NEEDLES OF TWENTY-EGHT *PICEA* SPECIES

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**Key Word Index**—*Picea*; Pinaceae; estolide waxes; chemotaxonomy; *n*-alkanes;  $\omega$ -hydroxyalkanoic acids; GC-MS.

**Abstract**—The needle wax of twenty-eight species of *Picea* has been investigated. The quantitative patterns of the *n*-alkanes and  $\omega$ -hydroxyalkanoic acids isolated from these waxes support the view that the genus should not be divided into sections.

### INTRODUCTION

The epicuticular wax of conifer needles contains normal monomeric wax esters [1], in addition to the characteristic polymeric esters, the estolides [2], formed from a series of  $\omega$ -hydroxyalkanoic acids either by self-esterification or by esterification with long-chain diols [3]. The quantitative variation in the individual  $\omega$ -hydroxy acids from these estolides have been successfully used as taxonomic discriminants in the genus *Pinus* (Pinaceae) and in the Cupressaceae [4]. The composition of the wax from *Picea* (Pinaceae) has been investigated to a limited extent [5-7] but no previous attempt has been made to use the wax components as aids in the taxonomy of this genus.

### RESULTS AND DISCUSSION

In the present study, the composition of the wax from needles of twenty-eight species of *Picea* was investigated. Each species yielded two types of wax: non-estolide wax which was extracted with *n*-hexane, and the chloroform-soluble estolide wax. The estolide wax was saponified to yield the individual  $\omega$ -hydroxyalkanoic acids. These were identified by GC-MS of their acetoxymethyl esters. The results of the quantitative analysis of the  $\omega$ -hydroxy acids and of the *n*-alkanes are given in Tables 1 and 2 respectively. The species are listed in order of their most abundant homologue. The  $\omega$ -hydroxy acid patterns closely resemble those obtained for the Cupressaceae and Pinaceae [4]. The proportion of even-numbered alkanes is high and there is no obvious dominant homologue. A similar situation was noted with *Pinus* species, whereas in *Cupressus* species the  $C_{33}$  alkane was strikingly dominant.

It was not possible to ensure that the composition of the wax from each of the twenty-eight species did not represent an extreme of intraspecific variation. However, a pilot study using two trees from each of three species showed little variation in the percentage  $\omega$ -hydroxyalkanoic acids: *Picea bicolor*  $C_{12}$  = 22%;  $C_{14}$  = 29%;

$C_{16}$  = 46%. *P. koyami*,  $C_{12}$  = 23%;  $C_{14}$  = 34%;  $C_{16}$  = 43%. *P. likiangensis*  $C_{12}$  = 67%;  $C_{14}$  = 16%;  $C_{16}$  = 17%. A variation of % hydroxy acids from the geographical standpoint is negligible. For example the values for *P. pungens* (Table 1) compare favourably with values reported for the species from Canada [6] and Kew [4].

The published taxonomic treatments of *Picea* were subjected to a process of evaluation by comparing them with both the  $\omega$ -hydroxy acid patterns (Table 1) and the *n*-alkane composition (Table 2). Some authorities divide the genus into two [8-12] and others into three sections [13-17]. Wright [18] studied morphological, distributional and genetic data for *Picea* and could find no support for such sectional divisions; the present results agree with this conclusion. Our data however support the view of Wright [18] that *P. obovata* is intermediate between *P. abies* and *P. asperata* and also that that taxa referred to as *P. asperata*, *P. gemmata* and *P. meyeri* all belong to the same species complex [14]. In addition, the fact that *P. balfouriana* and *P. likiangensis* have similar hydroxy acid composition agrees with morphological evidence indicating that these two taxa are either varieties [16] of the same species or two very closely related species [18].

### EXPERIMENTAL

Needles were collected in early May from named collections in the National Botanic Gardens, Dublin. Needles were collected from three levels (30 cm, 1 m and 2 m) and from four different aspects of each tree and pooled. Needles from the current years growth were rejected.

**Extraction of needle wax.** The air-dried needles (40-100 g) for each species were dipped in *n*-hexane (400 ml) for 1 min, dried in a current of air, and then dipped in  $CHCl_3$  (400 ml) for 1 min. The *n*-hexane and  $CHCl_3$  extracts were evaporated *in vacuo*.

**Analysis of *n*-alkanes.** *n*-Alkanes were obtained from the *n*-hexane extract by prep TLC (Si gel/*n*-hexane). The individual alkanes from each species were identified by GLC (3% SE-30 on Chromosorb G. AW. DMCS.  $N_2$  at 25 ml/min. temp. 230°) by comparison of their retention data with those of standard

Table 1. Mole percentage of  $\omega$ -hydroxyalkanoic acids in the needle wax of *Picea* species

<i>Picea</i> species	Classification of Pilger [10]	Classification of Harrison [16]	$\omega$ -Hydroxyalkanoic acids			
			C <sub>12</sub>	C <sub>14</sub>	C <sub>15</sub>	C <sub>16</sub>
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C <sub>12</sub> Dominant	Section					
<i>P. balfouriana</i> Rehder & Wilson	Eupicea	—	54	11	5	28
<i>P. likiangensis</i> (Franch.) Pritzel	Eupicea	Casicta	57	17	2	24
<i>P. morrisonicola</i> Hay.	Eupicea	Eupicea	36	19	10	34
<i>P. yezoensis</i> (Sieb et Zucc.) Carr.	Omorica	Casicta	35	27	13	25
<i>P. schrenkiana</i> Fisch. & Meyer	Eupicea	Eupicea	39	24	17	20
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C <sub>14</sub> Dominant						
<i>P. abies</i> (L.) Karst.		Eupicea	20	60	—	20
<i>P. breweriana</i> S. Wats.	Omorica	Omorika	20	37	10	33
<i>P. glauca</i> (Moench) Voss.	Eupicea	Eupicea	18	51	—	31
<i>P. omorika</i> Purkyně	Omorica	Omorika	27	35	24	14
<i>P. pungens</i> Engelm.	Eupicea	Casicta	22	54	—	24
<i>P. sitchensis</i> (Bong.) Carr	Omorica	Casicta	13	56	—	31
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C <sub>16</sub> Dominant						
<i>P. bicolor</i> (Maxim.) Mayr	Eupicea	Eupicea	18	34	—	48
<i>P. gemmata</i> Rehder & Wilson	Eupicea	—	36	18	5	41
<i>P. koyamai</i> Shiras.	Eupicea	Eupicea	25	33	—	42
<i>P. mariana</i> (Mill.) B.S.P.	Eupicea	Eupicea	5	45	—	50
<i>P. meyeri</i> Rehder & Wilson	Eupicea	—	20	22	18	40
<i>P. montigena</i> Masters	Eupicea	—	23	25	6	46
<i>P. obovata</i> Ledeb.	—	Eupicea	26	34	—	40
<i>P. orientalis</i> (L.) Link.	—	Eupicea	1	13	2	84
<i>P. purpurea</i> Masters	Eupicea	—	35	21	—	44
<i>P. sargentiana</i> Rehder & Wilson	Omorica	—	19	37	—	44
<i>P. smithiana</i> (Lamb.) Boiss.	Eupicea	Eupicea	9	15	3	73
<i>P. spinulosa</i> (Griff.) Henry	Omorica	Omorika	15	23	—	62
<i>P. wilsonii</i> Masters	Eupicea	Eupicea	29	29	—	42
<i>P. asperata</i> Masters	Eupicea	Eupicea	40	17	—	43
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No acid dominant						
<i>P. maximowiczii</i> (Regel) Masters	Eupicea	Eupicea	35	35	—	30
<i>P. brachytyla</i> (Franch.) Pritzel	Omorica	Omorika	39	21	—	40
<i>P. engelmannii</i> Parry ex Engelm.	Eupicea	Casicta	30	22	18	30

\* (—) not recognized in this classification.

alkanes and from the straight line plot of log *Tr* vs hydrocarbon number [19]. The relative amount of each alkane was determined from digital integrator data.

**Analysis of  $\omega$ -hydroxyalkanoic acids.** The  $\text{CHCl}_3$  extract for each species was saponified with ethanolic KOH (2N) at 100° for 3 hr. Non-saponifiables were removed with *n*-hexane. The fatty acids were removed by acidification (HCl) and extraction with  $\text{Et}_2\text{O}$ . The fatty acid mixtures were methylated and the methyl esters removed by extraction with  $\text{Et}_2\text{O}$ . The methyl  $\omega$ -hydroxyalkanoates were obtained by PLC (Si gel/ $\text{CHCl}_3$ - $\text{EtOAc}$ , 7:3). The band (*ca*  $R_f$  0.58) when eluted with  $\text{CHCl}_3$  and conc co-chromatographed (Si gel,  $\text{CHCl}_3$ - $\text{EtOAc}$ , 7:3) with a sample of authentic  $\omega$ -16-methylhydroxyhexadecanoate. The  $\omega$ -methylhydroxyalkanoates were acetylated ( $\text{Ac}_2\text{O}$ -Py) and the resulting products co-chromatographed (Si gel,  $\text{CHCl}_3$ -*n*-hexane, 3:1) with authentic  $\omega$ -16-methylacetoxylhexadecanoate.

**Identification.** The individual  $\omega$ -methylacetoxylalkanoates were identified by a comparison of the retention data with those of standard compounds (3% SE 30. on chromosorb GAW. DMCS.  $\text{N}_2$  at 25 ml/min temp. 205°). The identifications were confirmed by GC-MS (Finnigan 3200F series GC-MS system under the control of a 6110 data system; conditions: OV-1, 3%, temp. 180–240° at 2°/min, ionizing voltage 70 eV mass range 35–400 AMU.). The relative amount of each methylacetoxyl

alkanoate in the mixture from each species was determined from digital electronic integrator data.

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## REFERENCES

1. Kariyone, T., Takahashi, M., Watanabe, K., Ageta, H., Isoi, H., Hsu, H. Y., Kawano, N., Sawala, T. and Fukui, Y. (1962) *J. Pharmacog. Soc. Japan* **16**, 1.
2. Bougault, J. and Bourdier, L. (1909) *J. Pharm. Chim.* **29**, 561.
3. Kariyone, T., Isoi, K. and Yashikura, M. (1959) *J. Pharm. Soc. Japan* **79**, 61.
4. Herbin, G. A. and Robins, P. A. (1968) *Phytochemistry* **7**, 1325.

Table 2. Mole percentages of *n*-alkanes in the needle wax of *Picea* species

<i>Picea</i> species	Classification of		<i>n</i> -Alkanes																
	Pilger [10]	Harrison [16]	C <sub>19</sub>	C <sub>20</sub>	C <sub>21</sub>	C <sub>22</sub>	C <sub>23</sub>	C <sub>24</sub>	C <sub>25</sub>	C <sub>26</sub>	C <sub>27</sub>	C <sub>28</sub>	C <sub>29</sub>	C <sub>30</sub>	C <sub>31</sub>	C <sub>32</sub>	C <sub>33</sub>	C <sub>34</sub>	C <sub>35</sub>
<i>P. morrisonicola</i> Hay.	Eupicea	Eupicea	5	8	12	11	12	5	12	8	4	2	2	1	3	2	7	—	6
<i>P. bicolor</i> (maxim.) Mayr.	Eupicea	Eupicea	6	8	8	3	6	3	9	2	15	7	8	3	6	2	8	—	4
<i>P. breweriana</i> S. Wats.	Omorica	Omorica	9	8	12	4	13	T*	11	T	14	7	7	2	5	1	5	T	T
<i>P. gemmata</i> Rehder & Wilson	Eupicea	—	3	2	T	T	8	T	3	T	39	9	13	5	5	6	6	—	T
<i>P. koyamai</i> Shiras.	Eupicea	Eupicea	4	4	5	5	10	T	6	5	18	3	13	4	13	5	6	T	1
<i>P. likiangensis</i> (Franch.) Pritzel	Eupicea	Casicta	3	3	3	3	6	T	7	—	4	5	13	7	13	6	11	1	3
<i>P. montigena</i> Masters	Eupicea	—†	3	4	2	T	6	T	5	2	16	7	12	7	9	7	13	T	3
<i>P. obovata</i> Ledeb.	—†	Eupicea	6	7	7	2	4	T	7	1	16	6	10	3	13	3	15	—	—
<i>P. sargentiana</i> Rehder & Wilson	Omorica	—	3	3	4	5	1	T	10	T	36	4	9	1	8	2	12	—	T
<i>P. yezoensis</i> (Sieb et Zucc.) Carr.	Omorica	Casicta	5	3	6	4	7	2	5	2	14	6	10	6	12	5	11	—	1
<i>P. brachytyla</i> (Franch.) Pritzel	Omorica	Omorica	6	7	12	2	10	2	7	T	9	4	15	7	5	1	8	—	—
<i>P. engelmannii</i> Parry ex Engelm.	Eupicea	Casicta	1	3	4	—	3	—	1	—	8	11	17	8	13	6	16	—	T
<i>P. purpurea</i> Masters	Eupicea	—	T	T	2	2	4	T	5	T	9	5	16	8	15	10	15	—	—
<i>P. schrenkiana</i> Fisch. & Meyer	Eupicea	Eupicea	8	7	8	4	9	3	7	5	9	7	11	5	6	1	7	—	2
<i>P. spinulosa</i> (Griff.) Henry	Omorica	Omorica	5	8	9	T	8	1	11	2	14	8	14	2	5	1	7	—	T
<i>P. maximowiczii</i> (Regel) Masters	Eupicea	Eupicea	3	4	7	5	5	3	7	2	8	5	8	4	12	2	11	—	5
<i>P. wilsonii</i> Masters	Eupicea	Eupicea	4	6	7	2	4	T	5	1	12	8	13	6	15	7	9	—	T
<i>P. abies</i> (L.) Karst.	—	Eupicea	T	T	2	2	2	1	3	2	6	4	14	7	15	7	26	T	8
<i>P. asperata</i> Masters	Eupicea	Eupicea	8	9	6	4	10	T	9	T	12	5	5	2	8	2	13	1	1
<i>P. balfouriana</i> Rehder & Wilson	Eupicea	—	3	4	5	3	7	T	6	T	12	6	10	6	13	5	18	—	T
<i>P. glauca</i> (Moench) Voss.	Eupicea	Eupicea	3	2	3	2	3	T	10	T	14	10	15	3	8	3	16	—	3
<i>P. mariana</i> (Mill.) B.S.P.	Eupicea	Eupicea	1	1	2	2	3	2	4	2	9	3	10	6	10	6	26	3	8
<i>P. meyeri</i> Rehder & Wilson	Eupicea	—	1	2	1	2	4	T	4	T	11	5	13	6	19	6	22	T	2
<i>P. omorica</i> Purkyně	Omorica	Omorica	7	6	8	7	9	—	9	—	9	4	8	T	5	7	17	—	—
<i>P. orientalis</i> (L.) Link.	—	Eupicea	5	4	5	7	6	—	4	3	11	6	11	3	9	2	20	—	—
<i>P. pungens</i> Engelm.	Eupicea	Casicta	2	3	3	3	2	2	2	T	6	3	13	13	14	14	18	—	T
<i>P. sitchensis</i> (Bong.) Carr.	Omorica	Casicta	5	2	4	1	4	T	2	2	3	3	12	10	14	10	19	—	7
<i>P. smithiana</i> (Lamb.) Boiss.	Eupicea	Eupicea	2	3	4	4	2	1	4	1	7	8	11	9	9	9	14	2	8

\* T trace less than 1%. † Not recognized in this classification.

5. Stransky, K., Streibl, M. and Herout, V. (1967) *Coll. Czech. Chem. Commun.* **32**, 3213.
6. von Rudloff, E. (1959) *Can. J. Chem.* **37**, 1038.
7. Holloway, P. J., Jeffree, C. E. and Baker, E. A. (1976) *Phytochemistry* **15**, 1768.
8. Kent, A. H. (1900) *Veitch's Manual of the Coniferae* p. 424. Veitch, London.
9. Beissner, L. (1909) *Handbuch der Nadel Holz-Kunder*. Parey, Berlin.
10. Pilger, R. (1926) *Die Natürlichen Pflanzenfamilien* Vol. 13, p. 321. Englemann, Leipzig.
11. Florin, R. (1931) *Kungl. Svenska. Vetenskapsakad Hand.* **10**, 321.
12. Pilger, R. and Melchior, H. (1954) in *Syllabus der Pflanzenfamilien* (Melchior, H. and Werdermann, E. eds) 12th edn, Vol. 1, p. 330. Gebrüder Borntraeger, Berlin.
13. Mayr, H. (1890) *Monographie der Abietineen des Japanischen Reiches*. München.
14. Rehder, A. (1947) *Manual of Cultivated Trees and Shrubs* 2nd edn, p. 23. MacMillan, New York.
15. Fitzpatrick, H. M. (1965) *Scientific Proceedings of the Royal Dublin Society* p. 67.
16. Dallimore, W. and Jackson, A. B. (1966) *Handbook of Coniferae and Ginkgoaceae* (revised by Harrison, S. G.) 3rd edn. Edward Arnold, London.
17. Gaussen, M. (1966) *Les Gymnospermes* Fascicule VIII. Toulouse Faculté des Sciences.
18. Wright, J. W. (1955) *Forest Sci.* **1**, 319.
19. James, A. T. (1960) *Methods of Biochemical Analysis* (D. Gluck ed.) Vol. 8, 1 Interscience, New York.